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Abstract: The incorporation rate of Zn into the calcareous tests of *Pseudotriloculina rotunda* was investigated in culture in order to evaluate the possibility to use Zn/Ca ratios as pollution proxy. Foraminifera were incubated at zinc concentrations up to 10-fold higher than unpolluted seawater (sea+10 mg Zn/L) during 70 days. New calcite was observed at Environmental Scanning Electron Microscopy (ESEM), for potential alteration of test structure. Laser ablation-Inductively Coupled Plasma-Mass spectrometer (LA-ICP-MS) was used to quantify Zn contents. The analyses revealed that test structure is not visibly altered by the presence of zinc. However, significant Zn incorporation is detected by LA-ICP-MS. The zinc partition coefficient, D_{Zn} , decreases at increasing Zn concentrations (from 4.03 ± 0.06 at control conditions to 0.2 ± 0.01 at the highest tested Zn concentration) and the zinc is incorporation into the calcite follows a power function.

1 **Zinc incorporation in the miliolid foraminifer *Pseudotriloculina rotunda* under laboratory**
2 **conditions**

3

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14

15 **Highlights**

16 1- Zinc incorporation into miliolid foraminiferal tests was experimentally tested; 2- *Pseudotriloculina*
17 *rotunda* was cultured at several zinc concentrations for 70 days; 3- LA-ICP-MS analyses show that Zn
18 is incorporated into calcite; 4- Partition coefficients (D_{Zn}) decrease at increasing Zn/Ca_{seawater} ratios.

19

20

ABSTRACT

21 The incorporation rate of Zn into the calcareous tests of *Pseudotriloculina rotunda* was investigated
22 in culture in order to evaluate the possibility of using Zn/Ca ratios as a pollution proxy. Foraminifera
23 were incubated at zinc concentrations up to 10-fold higher than unpolluted seawater (sea+10 mg
24 Zn/L) during 70 days. New calcite was investigated under Environmental Scanning Electron
25 Microscope (ESEM), for potential alteration of test structure. Laser ablation-Inductively Coupled
26 Plasma-Mass spectrometry (LA-ICP-MS) was used to quantify Zn contents. The analyses revealed that
27 test structure is not visibly altered by the presence of zinc. However, significant Zn incorporation is
28 detected by the LA-ICP-MS. The zinc partition coefficient, D_{Zn} , decreases at increasing Zn
29 concentrations (from 4.03 ± 0.06 to 0.2 ± 0.01) and the zinc is incorporated into the calcite, non-
30 necessarily linearly.

31

32

KEYWORDS

33 Foraminifera; Miliolid; Zn/Ca; LA-ICP-MS; culture experiments; pollution.

34

35 **1. Introduction**

36 The recent worldwide legislation aims to restore the “pre-anthropic impacts” status in marine
37 environments (e.g., WFD, 2000/60/EC and MSFD, 2008/56/EC in Europe). Information about the
38 pristine faunas like those in pre-industrial times, however, are often impossible to obtain because of
39 the scarcity (or lack) of reference stations that could still represent unimpacted present-day

40 conditions. Fossilizing organisms represent an excellent historical archive of environmental
41 conditions. Foraminifera are distributed worldwide in many different habitats, from brackish to
42 marine, and their fossil records create an excellent historical archive which can be used as proxies for
43 the reconstruction of past environments, such as pre-industrial ecological conditions (Schönfeld et al.
44 2012). A new approach involves the use of foraminiferal test geochemistry to assess the evolution of
45 pollutant (i.e., metals) concentrations through time. Incorporation rates of trace elements are widely
46 used as specific proxies in paleoceanography and paleoecology (e.g., Eggins et al. 2003; Hönisch and
47 Hemming 2005; Levi et al. 2007; Katz et al. 2010; Sabbatini et al. 2011), despite the possible bias
48 linked to the biological influence on calcification processes (i.e., vital effects). The need to calibrate
49 these proxies through culturing experiments was highlighted in the last decade by several authors
50 (e.g., de Nooijer et al. 2007). This approach offers the advantage of changing one single variable
51 (while all the others are kept constant) in order to better evaluate the vital effect. These biological
52 aspects could be even more important for the incorporation rates of chemicals whose concentrations
53 exceed natural baselines due to human activity, and that could be potentially used as pollution
54 markers.

55 In this study we investigate the incorporation rates of Zn in the shell of the benthic miliolid
56 foraminifer *Pseudotriloculina rotunda* (Schlumberger 1893). Among foraminiferal species miliolids
57 showed contradictory responses to heavy metal pollution in different studies. For example,
58 decreasing miliolid relative abundances in polluted (by both organic and inorganic chemicals) coastal
59 zones are reported and interpreted by some authors as a sensitivity index (e.g., Ferraro et al. 2006;
60 Frontalini and Coccioni 2008). Other studies, on the other hand, suggest a strong tolerance of several
61 miliolid species to pollution, both *in situ* and under laboratory conditions (e.g., Samir and El-Din
62 2001; Romano et al. 2008; Cherchi et al. 2009; Foster et al. 2012; Nardelli et al. 2013).

63 The aim of the present study is to calibrate incorporation rates of Zn in miliolid foraminiferal shells
64 and thus evaluate the usefulness of the Zn incorporation rate as an environmental proxy. Zn is, in
65 fact, one of the most common pollutants associated with human activities (e.g., Callender and Rice

66 2000; Wuana and Okieimen 2011), that can be toxic for biological systems when its concentration
67 exceeds a threshold value (e.g., Haase et al. 2001; Valko et al. 2005; Díaz et al. 2006; Formigari et al.
68 2007). Nardelli et al. (2013) showed that inorganic Zn at concentrations higher than 0.1 mg/L can
69 cause biological stress in *Pseudotriloculina rotunda*, causing delay in calcification rates.

70 In this regard, our study also aims to test the hypothesis that the biological stress caused by high Zn
71 concentrations may influence metal incorporation rates as well. Zn incorporation was investigated
72 using the Laser Ablation Inductively Coupled Plasma Mass Spectrometer (LA-ICP-MS) analysis.
73 Moreover, morphological observation using the Environmental Scanning Electron Microscope (ESEM)
74 was employed to check for abnormalities in the organization/distribution of single shell crystallites.
75 Nardelli et al. (2013) previously suggested that Zn does not cause macroscopic test deformations in
76 miliolid foraminiferal shells, on the base of their observation of coiling patterns and chamber shapes
77 of *P. rotunda* specimens grown at increasing zinc concentrations, under the binocular microscope.
78 The aim of our ESEM analyses was to deepen these observations and check for calcite anomalies at
79 the crystallite level.

80

81 **2. Material and Methods**

82 *2.1 Experimental set up*

83 All the analyses were performed on specimens of *Pseudotriloculina rotunda* that grew at least one
84 chamber during 70-days exposure to six different Zn concentrations under laboratory conditions. Zn-
85 enriched solutions were prepared adding respectively 0.01, 0.1, 1.0, 10, and 100 mg/L of Zn to
86 natural seawater (sea) from an unpolluted site (Portonovo, Adriatic Sea); Zn and Ca concentrations in
87 natural seawater were measured using Inductively Coupled Plasma Mass Spectrometer (ICP-MS).
88 Although precautions were taken during manipulations, the ICP-MS analyses revealed that the zinc
89 concentration of the natural seawater used for the experiment was higher than natural background.
90 In fact the zinc concentration measured on culture waters before the addition of zinc was 0.149 ± 0.01
91 mg/L, while the zinc concentration of seawater from the sampling site was originally of 0.237 ± 0.01

92 $\mu\text{g/L}$. However, as the same water was used to prepare all the zinc solutions for the different
93 treatments and, considering the very high concentrations of added zinc, we believe that this
94 contamination does not compromise the dataset. But, of course, this means that the lowest tested
95 seawater zinc concentration of the experiment (named “sea” hereafter) cannot be considered as a
96 control representative of unpolluted seawater conditions.

97 Temperature ($15.0\pm 0.5^\circ\text{C}$), salinity (38.0 ± 0.001) and pH (8.0 ± 0.1) were kept constant during the
98 experiment. Refer to Nardelli et al. (2013) for further details on culture settings and preparation of
99 Zn solutions.

100 As reported in Nardelli et al. (2013), none of the specimens produced new chambers at the highest
101 tested Zn concentration (sea+100 mg/L), therefore only specimens coming from culture sets from
102 treatments sea, sea+0.01, sea+0.1, sea+1 and sea+10 mg/L were investigated. Moreover, two
103 samples were treated for a “passive Zn incorporation test”: empty tests of *P. rotunda* were exposed
104 to sea+10 mg/L Zn concentrations for two weeks in order to measure Zn passively adsorbed to
105 calcite, without involving cellular-mediated mineralization processes.

106

107 *2.2 ESEM and LA-ICP-MS sample preparation*

108 The analyzed foraminiferal tests ($n=41$) were washed with millipore water and dried at 40°C . The
109 same cleaning procedure was performed on the two empty tests ($n=2$) used for the passive
110 incorporation test. To perform both ESEM observations and LA-ICP-MS analyses, samples were fixed
111 to aluminium stubs using conductive carbon adhesive discs. All samples analysed with an LA-ICP-MS
112 were photographed under the ESEM before and after the analyses to check for the success of
113 ablations (i.e., the correct chamber, no multiple chamber sampling, and no breakage of chambers –
114 see Appendix, Figs. A.1a-c).

115

116 *2.3 LA-ICP MS analysis: analytical protocol optimization*

117 This study represents the first LA-ICP-MS investigation on miliolid foraminifera. Chemical
118 composition, micro-structure and chamber arrangements of miliolids strongly differ from other
119 benthic foraminifera more commonly analyzed with the LA-ICP-MS (i.e., Rotaliidae, e.g., de Nooijer et
120 al. 2007; Munsel et al. 2010; Dissard et al. 2010; or Buliminidae, e.g., Hintz et al. 2006; Barras et al.
121 2010). Miliolid foraminifera have a calcareous non-lamellar imperforate test consisting of calcite
122 needles randomly oriented in an organic matrix. They also possess a smoothly finished outermost
123 layer of well crystallized calcite, with rhombohedral crystal faces arranged parallel to the surface
124 (Debenay et al. 1998). Moreover, *Pseudotriloculina rotunda* creates chambers, each one-half coil in
125 length, adding the new ones in planes oriented at 120°, with only three final externally visible
126 chambers (Loeblich and Tappan 1964).

127 For this reason it was necessary to optimize the existing protocols and to obtain the best ablation
128 setting to be applied to our specimens. In particular, it was compulsory to prevent the laser from
129 ablating the innermost (older) chambers. Several trials on foraminiferal tests were thus performed to
130 find the most suitable combination of the instrument setting parameters. A detailed description of
131 the followed procedures is given in Appendix A, and the values of the optimized parameter used for
132 measurements are reported in Table 1. An example of successful sampling is shown in Figure 1.

133

134 *2.3.1 Analytical standards preparation and instrument calibration*

135 A mass spectrometer, like any measurement device, requires a suitable calibration procedure. When
136 laser ablation is employed, the interaction between laser and solid sample is complex and the
137 response is dependent on the sample matrix. For this reason, two forms of calibration are
138 mandatory: i) a reference (internal standard) is required to compensate for changes in the quantity
139 of ablated mass, even when the concentration remains constant; ii) matrix-matched solid standards
140 (frequently referred to as “external standards”) are necessary to calibrate laser ablation processes
141 and the instrument response. In fact, a relative measure of ablated mass can be achieved by
142 simultaneously measuring emission from the analyte and a common matrix element (internal

143 standard). For absolute calibration of the LA-ICP-MS conditions, standards made of the same matrix
144 as the samples would be required, but are seldom available (e.g., Darke and Tyson 1994; Raith et al.
145 1996; Hathorne et al. 2003).

146 Previous studies on perforate calcareous species of foraminifera (or other organisms with calcareous
147 compounds) used NIST610-611 or NIST612-613 as external standards, together with internal
148 standards (generally ^{44}Ca isotope) and “in-house made standards” (e.g., Hathorne et al. 2003; Eggins
149 et al. 2003; Montagna et al. 2007; Rathmann and Kuhnert 2008; Munsel et al. 2010). The same
150 standards were also used in our study in order to test their possible employment also for miliolids.
151 In-house made standards were obtained as hereafter described. Stock solutions with defined Zn
152 concentration were prepared using Zn ICP-standard solutions [1mg/mL in 2 (vol.%) HNO_3] and
153 Millipore water. A proper amount of each solution was added to a mixture composed by 400 mg of
154 ultrapure CaCO_3 powder (particle size less than 1 micron) and cellulose; to prevent CaCO_3 dissolution,
155 the pH of each solution was adjusted to 7.5 ± 0.1 using an appropriate amount of ammonia solution.
156 Each suspension was mixed and homogenized in an agate mortar and then dried at 30°C for 12
157 hours. The resulting powder was then re-homogenized in the agate mortar and pressed at 12 tons
158 into tablets of 12 mm diameter. Such “standard tablets” at different Zn concentrations were then
159 checked via LA-ICP-MS using ablation lines to verify whether Zn distribution was homogeneous. As
160 shown in Figure 2, the spectra resulting from these analyses revealed a fairly smooth plateau,
161 confirming a homogeneous distribution of the element into the standard tablets.

162 According to the literature (see above), NIST standards could be used as well. However, the
163 difference in composition of the matrix (i.e., silica glass in NIST standards, Ca carbonate in miliolids)
164 does not match the second requirement mentioned above. However, several measurements using
165 NIST610 were performed in order to test the possibility to use a certified and easily acquired
166 analytical standard. In particular, the same laser parameters (i.e., the same ablation conditions) were
167 applied to both NIST610 and our samples (miliolids) but the response on the internal standard (^{44}Ca)
168 was not satisfactory (see next paragraph for more details).

169 In the light of these results, and also in agreement with Hathorne et al. (2003), only in house-made
170 Tablets were used as external standards for measurements here reported. In detail, nine in house
171 made standards were used with Zn concentrations ranging from 0 (blank, CaCO₃+cellulose+Millipore)
172 to 1050 ppm.

173

174 *2.4 LA-ICP-MS: analyses of samples and data elaboration*

175 Eleven specimens from “sea”, five from sea+0.01 mg/L, six from sea+0.10 mg/L, ten from sea+1.0
176 mg/L, nine from sea+10 mg/L, were analyzed by the LA-ICP-MS. Moreover, the two specimens for the
177 “passive sorption tests” were investigated as well. In few cases more than one ICP-MS analysis was
178 carried out on the same chamber, otherwise one linear ablation per chamber was generally realized.
179 Standards were ablated using exactly the same laser setting parameters used for foraminifera, and
180 their concentrations were regularly measured during sampling procedure, in order to correct
181 instrumental deviation.

182 ⁶⁶Zn and ⁶⁸Zn isotopes, and ⁴⁴Ca were measured. A mix of helium (95%) and hydrogen (5%) were used
183 as reaction gas and a collision and reaction cell (KED) was used to minimize spectral interferences.
184 Both for calibration and for sample measurements, Ca and Zn concentrations were calculated
185 integrating over a time-interval of 1.00E+05ms in the flattest region of each spectrum of each
186 individual ablation profile using PlasmaLab™ Software Package 2007.

187 For data analyses the relation between Zn/Ca ratios in calcite and seawater was observed. Partition
188 coefficients for Zn were calculated for each set of Zn enriched cultures following the formula $D_{Zn} =$
189 $(Zn/Ca_{\text{calcite}})/(Zn/Ca_{\text{seawater}})$.

190

191 **3. Results**

192 *3.1 ESEM observations*

193 Results from ESEM observations on structure and crystal organization of Zn-exposed foraminiferal
194 tests did not reveal any obvious anomalies. All specimens showed the typical structure of miliolid
195 tests (Figs. 3 a-d), characterized by organized crystals on the external test surface and disorganized
196 crystals in an organic matrix in the inner part of the wall (Hay et al. 1963; Towe and Cifelli 1967;
197 Haake 1971; Debenay et al. 1998).

198 ESEM observations were also performed to adjust laser ablation parameters (see Appendix A) and to
199 check samples for correct ablation after LA-ICP-MS measurements.

200

201 *3.2 LA-ICP-MS measurements*

202 The main results of the LA-ICP-MS analysis are given in Figure 4 (a,b). The Zn seawater concentrations
203 are given in Zn/Ca mmol/mol to facilitate the comparison with Zn/Ca ratios in the calcite. The
204 different treatments (i.e., Zn concentrations) are indicated by the colors. The LA-ICP-MS analysis
205 revealed that Zn/Ca ratio increases linearly with water concentrations at least up to sea+1.0 mg/L
206 (Fig. 4b). The Zn/Ca shell concentrations vary between 1.11 ± 0.02 mmol/mol for the “sea” treatment
207 and 1.77 ± 0.04 mmol/mol of the treatment sea+1.0 mg/L (Tab. 2; see also Tab. A.2 in Appendix A for
208 the complete dataset). The measurements obtained on calcite produced at the highest tested Zn
209 concentrations (sea+10 mg/L), however, with average Zn/Ca values of 3.81 ± 0.17 mmol/mol, suggest
210 the possibility that the linear trend, observed for seawater Zn concentrations lower than sea+1.0
211 mg/L, can turn towards a plateau for higher concentrations. Possible explanations of this trend are
212 discussed later.

213 The passive sorption test on empty shells incubated at sea+10 mg/L conditions showed very low Zn
214 concentrations ($Zn/Ca_{\text{calcite}} = 1.17 \pm 0.01$ mmol/mol), comparable to the ones of samples from the
215 “sea” treatment ($Zn/Ca_{\text{calcite}} = 1.12 \pm 0.02$ mmol/mol), and more than 3 times lower than living samples
216 from highest tested zinc concentrations (sea+10 mg/L) ($Zn/Ca_{\text{calcite}} = 3.83 \pm 0.17$ mmol/mol) (Fig. 4 and
217 Tab. A.2 of Appendix A).

218 The calculated Zn partition coefficients (D_{Zn}) for each Zn treatment (indicated by the colors) are given
219 in Figure 5a. The average D_{Zn} (\pm standard deviation) is given in numbers. Because of inhomogeneous
220 variance, the Kruskal-Wallis test and Mann-Whitney pairwise comparisons post-hoc test were
221 performed to determine whether the difference between partition coefficients was significant
222 among treatments (p -value < 0.001). The results showed that the Zn partition coefficients were all
223 significantly different among treatments. The obtained average values of D_{Zn} varied between
224 4.03 ± 0.06 at “sea” conditions and 0.2 ± 0.01 at sea+10 mg/l Zn concentration (Fig. 5) (total average
225 $D_{Zn} = 2.22 \pm 1.56$). The observed decrease of D_{Zn} at increasing seawater Zn concentration is well
226 described by a power function ($R^2 = 0.997$; p -value < 0.001 ; see Fig. 5a). However, due to the difficulty
227 to assess whether the results obtained for the highest tested concentration (sea+10 mg/L) are real or
228 affected by neglected precipitation of zinc oxides/hydroxides (see discussion paragraph 4.2) we
229 reported in fig. 5b only the D_{Zn} obtained at Zn treatments lower than this concentration. Even
230 without the last concentration, the data are still well described by a power function ($R^2 = 0.998$; p -
231 value < 0.001) and the regression equation slightly change from $y = 1.5552x^{-0.713}$ to $y = 1.4826x^{-0.775}$.

232

233 4. Discussion

234 4.1 ESEM structural observation of tests

235 Miliolids are often regarded as pollution sensitive organisms. Several studies on test deformity
236 induced by environmental contamination suggest that miliolids are more easily affected by test
237 deformities than other foraminifera. For example, Samir and El-Din (2001) found that the majority of
238 deformed tests collected in the El-Mex Bay (Egypt) were miliolid shells. Also Sharifi et al. (1991)
239 reported that deformed foraminiferal tests from the Southampton coastal area contained much
240 higher Cu and Zn concentrations than non-deformed specimens, suggesting again a responsibility of
241 these metals for test deformations. However, Nardelli et al. (2013) tested in laboratory conditions
242 the biological effects of several Zn concentrations on *P. rotunda* and, despite the fact that some

243 specimens calcified new chambers at Zn concentrations up to 10 mg/L, no obvious deformations due
244 to Zn exposure were observed, in terms of chamber arrangements or general shape of the
245 foraminiferal shells. Our ESEM observations confirm these results because the calcite produced
246 during Zn exposure showed the typical aspect and arrangement known for the species (as described
247 by Debenay et al. 1998). Then, Zn exposure does not appear to be, by itself, a cause of abnormal
248 calcification for *P. rotunda*, either at the macro or micro scale. A possible explanation for the fact that
249 Zn is often found in anomalous tests is its possible covariance with other pollutants (for example
250 other metals) or environmental parameters, which can be alone the real cause of test deformation or
251 have synergic behaviors to zinc and enhance its toxicity (as demonstrated, for example, for Co and Zn
252 by Bresson et al. 2013). In fact, even if Zn is one of the most cited heavy metals, potentially
253 responsible for test deformation, many other pollutants and anthropogenic events were related to
254 test deformities, e.g. Cd, Cr and Ti (Yanko et al. 1998), Cu (Le Cadre and Debenay 2006),
255 hydrocarbons and oil spills (e.g., Vénec-Peyré 1981; Vénec-Peyré et al. 2010), rapid changes of
256 salinity and hypersalinity (e.g., Eichler-Coelho et al. 1996; Sousa et al. 1997; Stouff et al. 1999; Geslin
257 et al. 2002). The absence of visible test anomalies can be coherent with the hypothesis that Zn
258 incorporation into biomineralized calcite may occur, as already observed on non-biogenic calcites
259 (e.g., Elzinga and Reeder 2000; Temmam et al. 2000), after substitution of Ca or Mg to form
260 isomorphic Zn carbonates, as suggested by Madkour and Ali (2009).

261

262 *4.2 Zinc incorporation*

263 Despite the absence of deformations or visible abnormalities in the specimens analyzed in this study,
264 high Zn/Ca contents were detected in the internal wall of the last chamber of the test by LA-ICP-MS
265 analysis. The incorporation rates of Zn into the calcite do not seem to be constant at increasing Zn
266 concentrations. Zn incorporation, in fact, linearly increases at lower concentrations (from sea to
267 sea+1.0 mg/L, Fig. 4b), but the measures obtained at Zn concentrations higher than sea+1.0 mg/L,
268 even if represented by the results from only one Zn treatment, suggest that at higher concentrations

269 (i.e., sea+10 mg/L) the trend determined at low concentrations (up to sea+1.0 mg/L) is no longer
270 respected (Fig. 4a). The decreasing Zn incorporation rate in the shell is reflected by the partition
271 coefficients that are lower for higher Zn seawater treatments (Fig. 5a,b).

272 Considering that we worked at high zinc concentrations we cannot avoid to take into account that
273 zinc oxides/hydroxides precipitation could have been important at the highest zinc concentration we
274 tested (sea+10 mg/L). In fact, if we roughly estimate the expected Zn^{2+} saturation based on the pH of
275 our cultures (8.0 ± 0.1) and neglect all potential influences of kinetics, we obtain a concentration of
276 saturation of $4.37E-05$ M, which is 3 times lower than the concentration of zinc we added to this
277 treatment (see Tab. A.2 in Appendix A). Zinc oxides/hydroxides precipitation could therefore at least
278 partially explain the decrease of Zn incorporation at sea+10 mg/L condition.

279 However, this is not the first time that a similar decreasing trend for metal incorporation is observed
280 on benthic foraminifera and this could be also partially due to biological reasons. For example
281 Munsel et al. (2010) report similar observations for incorporated Ni in *A. tepida's* calcite. The
282 incorporation of the metal is linear at low experimental concentrations and drops down when
283 foraminifera are exposed to Ni concentrations 20-fold higher than natural seawater. The authors
284 suggest that this could be due to biological effects. In fact, according to them, Ni at high
285 concentrations could have toxic effects on foraminiferal cells and inhibit calcification. Toxic potential
286 of some highly concentrated metals could trigger their cellular expulsion or blocking mechanism (for
287 example metallothioneins-mediated) and prevent their incorporation in proportion to their sea
288 water concentrations. This hypothesis is also valid for our study. Nardelli et al. (2013) reported
289 delayed growth rates (i.e. calcification) of *Pseudotriloculina rotunda* exposed to Zn concentrations
290 higher than sea+1.0 mg/l and ascribed the reduced calcification rate to biological effects of high zinc
291 concentrations on foraminiferal cell, invoking similar processes. Thus we think that the potential
292 biological influence on the Zn incorporation observed at sea+10 mg/l in the present study cannot be
293 excluded. Some other possible explanations are hypothesized by Mewes et al. (2014) who also
294 observed an exponential decreasing partition coefficient for Mg/Ca at increasing seawater metal

295 concentrations following a power regression for two rotaliid foraminifera (*Ammonia aomoriensis* and
296 *Amphistegina lessonii*): (a) presence of two different CaCO_3 layers, (b) involvement of two different
297 biomineralization pathways. The first hypothesis depends on the possibility, shown for some
298 foraminifera, to precipitate two different calcite phases, with two different Mg/Ca calcite ratios,
299 independently of a migration of the foraminifera through different chemical environments (in the
300 water column or in the sediment). This pathway was described by Bentov and Erez (2005) and
301 particularly for *A. lessonii* and *Orbulina universa* by Branson et al. (2013). However, as far as we
302 know, this biomineralization mechanism has not been described and does not fit with the existing
303 calcification models for miliolid foraminifera. In fact the existing biomineralization models for miliolid
304 foraminifera propose two kinds of calcification mechanisms: some miliolid foraminifera form bundles
305 composed of an array of oriented crystals. Each crystal is enveloped by organic material, and then
306 bundles are passed through the cell membrane by exocytosis. Other miliolids, instead, accrete pre-
307 formed crystals and matrix materials onto extracellular surfaces to form a loosely packed wall
308 structure in a “stack-of-bricks” process (e.g., Berthold 1976; Hemleben et al. 1986).

309 The second hypothesis considers the involvement of different biomineralization pathways of Mg
310 incorporation during calcification. This hypothesis derives from the biomineralization model
311 proposed by Nehrke et al. (2013) for rotaliids foraminifera, but there is no evidence that the same
312 processes occur into miliolid foraminifera. Therefore also this hypothesis does not seem to be valid
313 to explain our results. Thus, the oversaturation of the highest tested zinc solution and/or the
314 biological effect of potentially toxic zinc concentrations on the biomineralization process remain, to
315 us, the more realistic hypothesis to explain the low zinc incorporation at sea+10 mg/L treatment.

316 A part from the exponential decrease of partition coefficient, another interesting point highlighted by
317 our results is the apparent positive intercept of D_{Zn} ($\text{Zn}/\text{Ca}_{\text{calcite}}$ vs $\text{Zn}/\text{Ca}_{\text{seawater}}$) regression (Fig. 4b).
318 This result suggests a difference in Zn incorporation for this miliolid species compared to all the
319 rotaliid foraminifera for which D_{Zn} was previously estimated (e.g. Marchitto et al., 2000; Bryan and
320 Marchitto, 2010). In fact, despite the highly variable D_{Zn} estimated for the different species,

321 Marchitto et al. (2000) and Bryan and Marchitto (2010), report a Zn/Ca_{calcite} vs Zn/Ca_{seawater} regression
322 line passing from the origin of the axes for all the measured rotaliids foraminifera. Similar results,
323 showing a positive intercept on the y-axis, reported in Mewes et al. (2014) for Mg/Ca of two rotaliid
324 foraminifera (*Ammonia aomoriensis* and *Amphistegina lessonii*), were hypothesized by the authors to
325 be possibly due to purely chemical reasons: sorption of Mg^{2+} to mineral surfaces would be stronger
326 than sorption of Ca^{2+} (Mucci and Morse 1983) and then increased Mg^{2+} (adsorbed on calcite) would
327 locally increase Mg/Ca, and then explain the positive y-axis intercept, especially at low seawater
328 Mg/Ca. Due to its divalent nature and its ionic radii smaller than Ca^{2+} , this kind of mechanism is also
329 possible for Zn^{2+} (Elzinga and Reeder, 2000). In this regard, the results of our sorption test (fig. 4)
330 seems to suggest that, at least at the highest tested concentration (sea+10 mg/L), Zn sorption is
331 negligible compared to the fraction incorporated into the calcite of living foraminifera incubated at
332 the same Zn concentrations. This suggests that this kind of mechanism could have low influence on
333 Zn enrichment of calcite. However, the test was performed only at the highest Zn treatment and it is
334 not enough to draw any definitive conclusion about this aspect.

335 Another hypothesis, invoked by Langer et al. (2006) to explain the enriched Sr/Ca ratio in the calcite
336 of the coccolithophore *Emiliana huxleyi*, could also explain the Zn/Ca enrichment we observe for
337 *Pseudotriloculina rotunda*. Similarly to miliolid foraminifera, coccolithophores precipitate calcite into
338 specific intracellular vesicles. According to the authors, the intracellular calcite precipitation yields an
339 accumulation of Sr in the coccolith vesicle until Sr steady state is achieved. Then, the observed Sr/Ca
340 of the coccolith calcite would be an integral value that arises from the sum of Sr/Ca over the time
341 required for the formation of one coccolith. This is suggested to explain the enriched Sr/Ca ratios
342 found into the coccolithophore compared to theoretical ones expected by purely chemical
343 precipitation. According to us, and keeping into account the results of our sorption test, this
344 hypothesis seems more convincing to explain the apparent positive y-axis intercept of figure 4.

345

346 *4.3 Implications for reconstructing past seawater zinc concentrations*

347 As zinc is one of the most diffused pollutants related to the industrial activity (e.g., Callender and
348 Rice 2000; Wuana and Okieimen 2011) a proxy for its concentrations in historical times could
349 represent an important tool to study the evolution of marine ecosystems under anthropic stress. This
350 is particularly interesting in view of the animated debates on the possible boundary for the recently
351 proposed new epoch of the Anthropocene (e.g., Crutzen 2002; Zalasiewicz et al. 2011). The
352 accelerated increase of some industrial-derived pollutants is, in fact, one of the proposed points to
353 establish the start of this epoch. Our results suggest a possible reliable identification and
354 interpretation of one of these pollutants in the historical sediment record. In fact, except for the D_{Zn}
355 estimated for the sea+10 mg/L that could have been influenced by Zn oversaturation problems (as
356 discussed above), we are confident that the D_{Zn} estimated for *Pseudotriloculina rotunda* in this study
357 can be applied for past seawater concentrations reconstructions. The partition coefficients we
358 obtained are different from the ones previously estimated for other foraminiferal species (mainly
359 Rotaliids), but rely on the variability range of values found for different species of rotaliid
360 foraminifera (e.g. ~ 3 for *U. peregrina* to ~ 22 for *C. pachyderma* reported by Bryan and Marchitto,
361 2010) and they represent the Zn/Ca of the seawater where the calcite was biomineralized. Zn/Ca
362 contents into miliolid calcite could therefore allow reconstruction of concentrations of this metal in
363 the past seawater.

364 The obtained D_{Zn} for each tested zinc concentrations (fig. 5a) show statistically significant differences.
365 For this reason we would suggest not to average all the values but rather using the equations given in
366 figure 4b and 5b for Zn seawater reconstructions. For the highest tested zinc concentration, for
367 which the estimated D_{Zn} could have been highly driven by zinc oxides/hydroxides precipitation and
368 physiological cell problems due to potential toxic effects of zinc, our results are more difficult to
369 apply and need further investigation to be confirmed. For the use of this proxy in potentially
370 contaminated environments, the possible toxic effect of Zn at high concentrations on *P. rotunda*
371 could be an important boundary constraint. A possible solution to overcome this problem could be
372 the use of multiple species, with different thresholds of tolerance to zinc.

373 **5. Concluding remarks**

374 Zn/Ca ratios in calcite of *Pseudotriloculina rotunda* are linear function of Zn/Ca in seawater at lower
375 Zn/Ca seawater and appear to turn into a power function at Zn seawater concentrations higher than
376 sea+1.0 mg/L. Further studies are needed to quantify the possible influence of zinc
377 oxides/hydroxides precipitation on the results we obtained at this higher experimental condition.
378 However, even if we omit the last point from the dataset, the results show that Zn is incorporated in
379 equilibrium with seawater concentrations and that partition coefficients significantly decrease at
380 increasing seawater zinc concentrations following a power function. The result of passive sorption
381 test performed on dead specimens incubated in water with high Zn concentration suggests that most
382 of the zinc incorporation is mediated by the cell and that chemical passive sorption is putatively
383 negligible (Tab. A.2). These results suggest the suitability of Zn/Ca into foraminiferal calcite as an
384 environmental proxy. Zinc contents of calcite can be in fact considered representative of the
385 concentrations in seawater at the time of their calcification, and relatively independent of post-
386 mortem conditions occurring in the depositional area. The analysis of Zn/Ca contents into the calcite
387 could therefore allow the reconstruction of past zinc concentrations in seawater. Then, present-day
388 knowledge about tolerance limits of foraminifera (e.g., de Freitas Prazeres et al. 2011; Nardelli et al.
389 2013) and other organisms to Zn seawater concentrations would allow us to study potential
390 ecological effects induced by high concentrations of this metal in environmental systems in the past.
391 However further studies would be needed for a more precise calibration of Zn/Ca as a proxy. In
392 particular the study of potential synergic, additive or antagonistic effects of several chemicals and/or
393 of environmental variables (e.g., salinity, temperature, alkalinity..) on Zn incorporation should be
394 deeply investigated. Moreover, further calibration will be very useful in the future to fill the gap for
395 concentrations between 1 and 10 mg/L and between 0.1mg/L and natural unpolluted seawater zinc
396 concentrations (e.g., one order of magnitude lower).

397

398

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408

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535 **Tables**

536 Tab. 1 Optimized laser parameters. Linear ablation, carried out on the flatter part of the last chamber
 537 proceeding always from the centre to the aperture of the chamber, was preferred to spot ablation.
 538 See Hathorne et al. (2003) for comparison and SI-1 for further information. A pre-ablation step was
 539 introduced to clean the surface from potential contaminations before data acquisition.

540

	Laser Intensity (%)	Frequency (Hz)	Ablation line width (μm)	Duration (s)	Main results
Pre-ablation	10	1	55	230	Sampling only of external chamber, without crash; good signal intensity at the detector.
Ablation	50	4	55	230	

541

542

543 Tab. 2 Mean $\text{Zn}/\text{Ca}_{\text{seawater}}$, $\text{Zn}/\text{Ca}_{\text{calcite}}$ and D_{Zn} values (\pm standard deviations) for each Zn treatment.

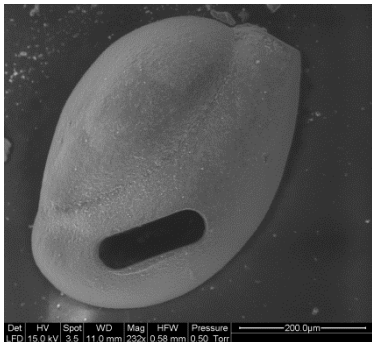
Treatment	$\text{Zn}/\text{Ca}_{\text{seawater}}$ (mmol/mol)	$\text{Zn}/\text{Ca}_{\text{calcite}}$ (mmol/mol) (\pm s.d.)	D_{Zn} (\pm s.d.)
sea (control)	0.278	1.122 (\pm 0.028)	4.034 (\pm 0.059)
sea+0.01 mg Zn/L	0.297	1.152 (\pm 0.012)	3.880 (\pm 0.042)
sea+0.1 mg Zn/L	0.465	1.216 (\pm 0.011)	2.616 (\pm 0.024)
sea+1.0 mg Zn/L	2.145	1.771 (\pm 0.037)	0.826 (\pm 0.017)
sea+10.0 mg Zn/L	18.948	3.813 (\pm 0.171)	0.201 (\pm 0.009)

544

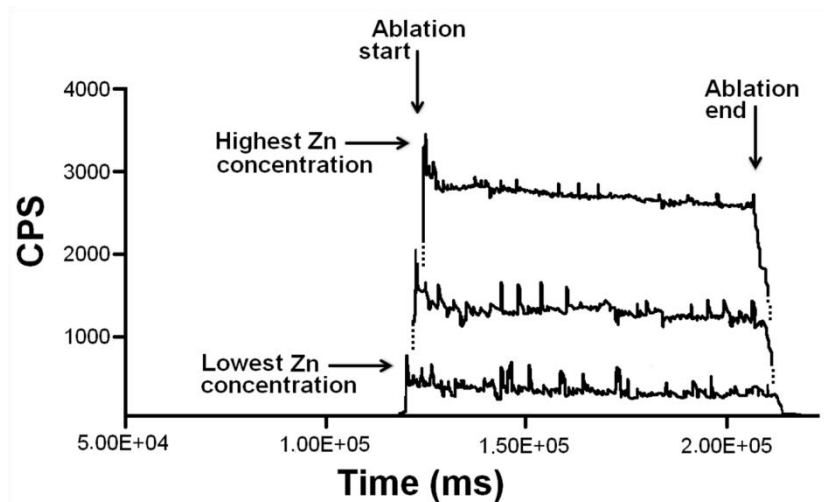
545

546 **Figures**

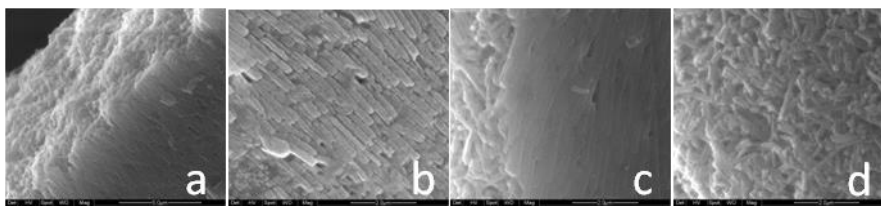
547 Fig. 1 Example of successful laser sampling on last chamber of *P. rotunda*



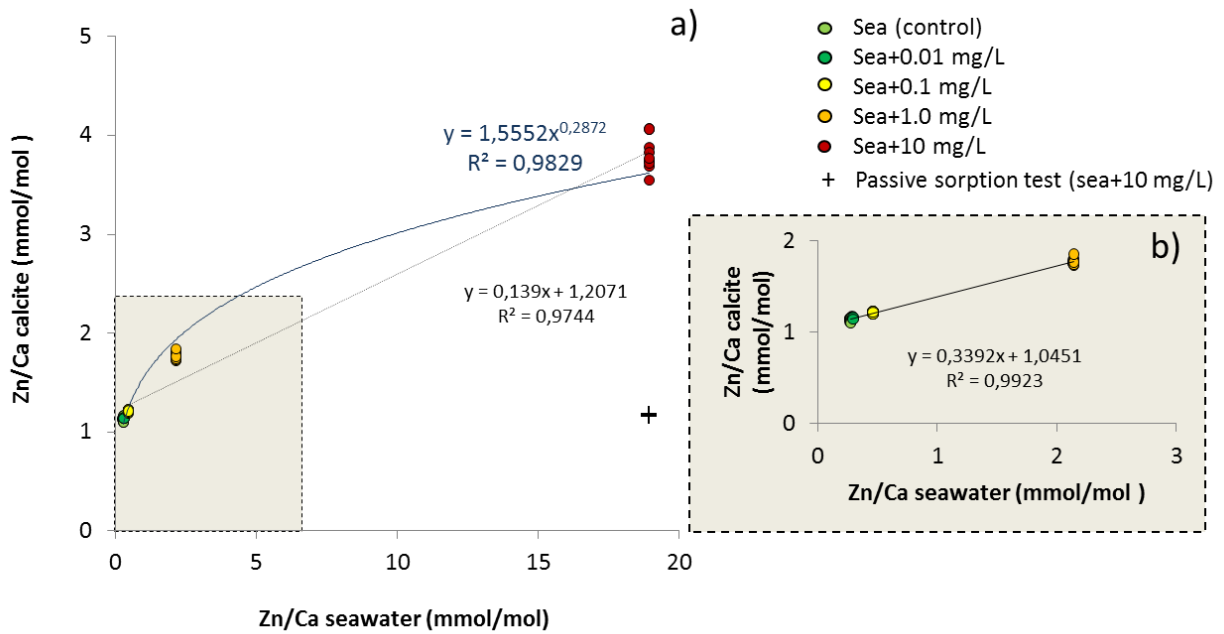
550 Fig. 2 Spectra resulting from the ablation lines measured on home-made standard tablets at different
551 Zn concentrations to verify if Zn distribution was homogeneous. The highest and the lowest curves
552 represent the spectra measured respectively on 1050 ppm and 150 ppm zinc home-made standards.
553 CPS means counting per second.



557 Fig. 3 ESEM micrographs of *P. rotunda*'s test. a) View of external wall of a specimen from control
558 cultures (sea); b) detail of crystals on the external wall of the same specimen; c) View of external wall
559 of a specimen from sea+10 mg Zn/L contaminated cultures and d) detail of internal part of the wall,
560 with disorganized crystals of the same specimen.

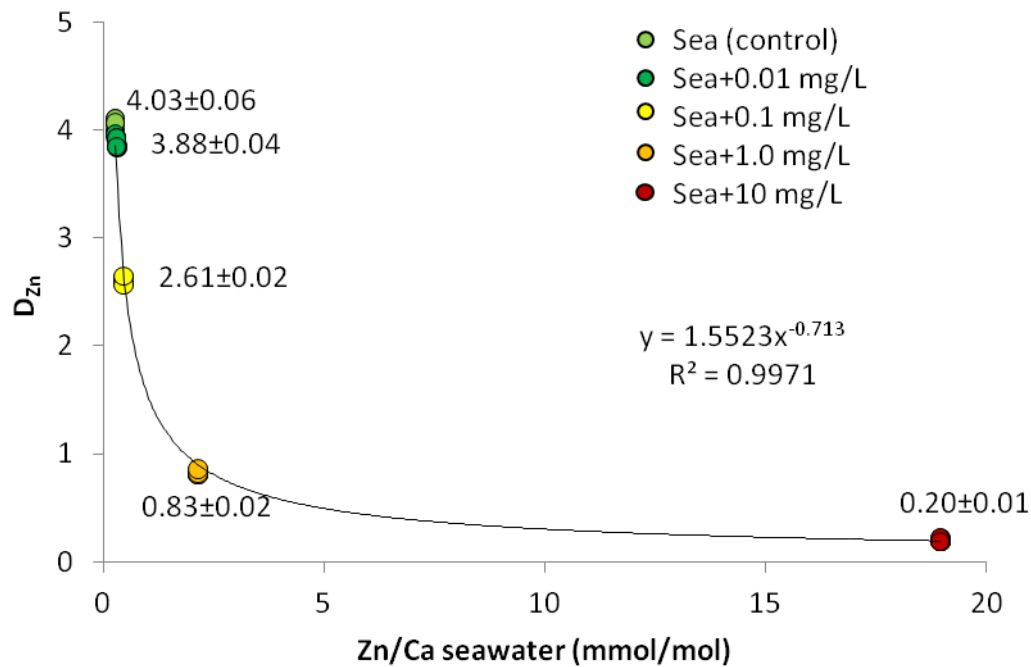


565 Fig. 4 Zn/Ca ratios in *Pseudotriloculina rotunda* calcite in function of Zn/Ca in seawater. a) All the
 566 samples are included (the colors correspond to the different Zn treatments). The best fitting of
 567 power function to predict $(Zn/Ca)_{calcite}$ to $(Zn/Ca)_{seawater}$ relationship is shown in blue and compared to
 568 the linear model (black dashed line). b) This is an enlargement of the left corner of figure 4a where
 569 the lower Zn/Ca points are showed separately.



570

571 Fig. 5 Partition coefficients measured for each zinc treatment. The averages $D_{Zn} \pm$ standard deviation
 572 for each treatment are given in numbers.



573

574

575

Supplementary Data for online publication only

[Click here to download Supplementary Data for online publication only: Appendix A.pdf](#)